

Therapeutic Potential of Secondary Metabolites from Endophytic Fungi in Biotechnology and Medicine

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Date Received: 06-10-2023 Date Accepted: 12-02-2024 DOI: https://doi.org/10.48198/NJPAS/23.B28

ABSTRACT

Endophytic fungi, which reside within plant tissues without causing harm, have attracted considerable scientific attention due to their wide-ranging applications. This review delves examines key aspects of research on endophytic fungi, focusing on isolation methods, the production of natural compounds, sampling strategies, and their therapeutic potential. Techniques such as surface sterilization and the use of selective media have revealed a vast diversity of fungi within plant hosts, shedding light on their ecological roles and biotechnological importance. Endophytic fungi are known to produce a variety of natural compounds, particularly secondary metabolites with medicinal properties. Prominent examples include taxol from *Taxus* endophytes and camptothecin from *Camptotheca acuminata* endophytes, both recognized for their anticancer activities. Various sampling strategies, from random collection to host-specific approaches, are essential for uncovering the diversity of these fungi. Additionally, optimization techniques for cultivation and extraction have been developed to maximize the yield of bioactive compounds and ensure sustainable production. Beyond anticancer agents, endophytic fungi have shown potential as sources of antimicrobials, antioxidants, and immunomodulators. These bioactive metabolites offer promising alternatives for drug development, particularly in response to the growing challenge of antibiotic resistance. This review contributes to the growing body of knowledge in the understanding of endophytic fungi and their applications across medicine, agriculture, and biotechnology.

Keywords: Antimicrobials, Bioactive compounds, Endophytic fungi. Secondary metabolites,

Introduction

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The prevalence of drug resistance is escalating rapidly, exacerbating the challenge of managing infectious diseases. The efficacy of existing antimicrobial drugs is increasingly compromised due to rising resistance cases, highlighting the urgent need for innovative solutions that offer lasting efficacy (Suleiman *et al.*, 2016; Pasrija *et al.*, 2022). In response to these health concerns, it is not surprising that many individuals, particularly

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in developing countries, are turning to naturally occurring bioactive compounds for primary healthcare (Anand *et al.*, 2019). Plants produce a wide array of secondary metabolites with significant medical, pharmacological, and industrial applications, all of which are vital to human well-being. However, the growing demand for these metabolites has endangered numerous medicinal plants. The processes involved in extracting plant secondary metabolites are often time-consuming, costly, and labor-intensive (Yeshi *et al.*, 2022).

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Plants form symbiotic associations with various microorganisms, leading to the term "endophytic," which has been widely discussed and defined in multiple ways. As first proposed by Anton De Bary in 1886, the term generally refers to any organism that resides within (éndon) a plant (phyton). Endophytic fungi (EF) represent a large and diverse group within endophytes, with an estimated one million different fungal taxa (Abdelaziz *et al.*, 2022). The endophytic community is further classified into two subgroups: 'obligate endophytes,' which depend critically on plant metabolism for survival, and 'facultative endophytes,' which can spend certain stages of their life cycle outside the host, often associating with plants in the surrounding soil environment (Jha et al., 2023).

Endophytes engage in a variety of interactions with their host plants, ranging from symbiotic and benign commensal relationships to decomposer and latent pathogenic associations (Diagra & Nonzom, 2023). Currently, the term "endophytic" encompasses a broad range of microorganisms, including bacteria, fungi, viruses, protozoa, and even microalgae, that inhabit plant tissues without causing harm to their hosts (Noemi & Everlon, 2022). Endophytic fungi can enhance host plant growth by releasing phytohormones or by increasing the plant's resilience to various stresses. Additionally, they can produce insecticides that protect plants from herbivores (Abdelaziz *et al.*, 2022). Under stress conditions, endophytes secrete a variety of stress-responsive molecules such as gibberellins (GB), cytokinins (CIS), salicylic acid (SA), and indole-3-acetic acid (IAA), which promote plant growth, development, and induce several physiological changes (Shaffique *et al.*, 2022).

Almost every plant, particularly woody species, hosts one or more fungal endophytes. These fungal endophytes are found in diverse geographical and climatic zones, displaying a wide distribution and high species diversity. Their ubiquity, diversity, and varied ecological roles suggest that they are well-adapted to their plant hosts under global distribution and selective pressure (Singh & Kumar, 2023). The identification of the plant as a host triggers the activation of virulence mechanisms necessary for colonization and induces host defense responses. This dynamic interaction, best described as balanced antagonism, allows the fungus to thrive by utilizing nutrients from the host while providing benefits such as resistance to biotic and abiotic stresses (Noemi & Everlon, 2022; Bamisile et al., 2018).

Endophytic fungi have become valuable resources in the pharmaceutical and healthcare sectors, serving as sources of novel therapeutic biomolecules. These fungi produce a variety of including immunosuppressants, compounds, anticancer agents, plant growth promoters, antimicrobial volatiles, insecticides, antioxidants, and antibiotics (Mamankey, 2024). Notable examples of medicinal endophytic fungi include Penicillium chrysogenum, Taxomyces andreanae, Piriformospora indica. and Curvularia protuberata. These fungi have been found to produce beneficial medicinal substances, such as paclitaxel (taxol) and camptothecin, both of which are anticancer agents, ergoflavin, which has antiinflammatory properties, and cyclosporine, an antiviral compound (Eid et al., 2021).

This review aims to explore the research surrounding endophytic fungi and their diverse medicinal potentials, with a focus on the isolation of endophytes, the natural production of bioactive compounds, and the optimization of sampling and media for the production of these bioactive molecules.

Isolation of Endophytic Fungi

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The concept of the "rhizosphere" was introduced by agronomist and plant physiologist Lorenz Hiltner in 1904, describing the area around a plant's root system that harbors a diverse microbial community. The term has since evolved, and the rhizosphere is now divided into three zones: the endo-rhizosphere, where microorganisms occupy spaces between plant cells; the rhizoplane, which is adjacent to the root epidermal cells; and the ectorhizosphere, which surrounds the roots and extends into the surrounding soil, housing free-living or non-symbiotic microorganisms (de Paula *et al.*, 2021).

One of the primary methods for isolating endophytic fungi involves the surface sterilization of plant tissues followed by culturing on selective media. This technique has been widely used across various plant species, revealing a rich diversity of endophytic fungi. For example, a significant number of endophytic fungi with diverse biotechnological potential have been isolated from the medicinal plant Bauhinia forficata (Bezerra et al., 2015). Similarly, Rajakumari et al. (2018) isolated 25 endophytic fungi from Calotropis procera, demonstrating their ability to produce various chemically significant compounds with pharmaceutical importance. The outcomes of the isolation process can vary depending on the type of plant tissue used; for example, in a study by Hastuti et al. (2021). Various studies have highlighted the diversity of endophytic fungi across different plant species and tissues. In one study, Hussaini et al. (2022) isolated 35 endophytic fungi from the leaves of Azadirachta indica and Mentha piperita,

identifying genera such as *Trichoderma*, *Penicillium*, and *Aspergillus* (Table 1). Other successful isolations from the leaves and stems of *Stenochlaena palustris* have shown that these fungi produce secondary metabolites with significant antibacterial properties (Aisyah *et al.*, 2023; Mashar *et al.*, 2023).

Natural Products as Bioactive Compounds

Natural products are substances biosynthesized by living organisms, including plants, microbes, and animals (Pang, 2023). The term most commonly refers to secondary metabolites, which are distinct from primary metabolites. While primary metabolites are directly involved in essential growth and developmental processes, secondary metabolites play indirect roles in metabolism and fulfill crucial ecological functions within the organism (Dadwal, 2024). The production of secondary metabolites by fungal endophytes is strongly influenced by various factors such as the timing of sample collection, environmental conditions, the geographical location or habitat of the host plant (with extreme environments like saline habitats, high altitudes, rainforests, deserts, and marshes being particularly swamps, significant), the nutritional source of the host plant, the specific plant tissues (such as roots, leaves, or seeds), and the type of plant (whether angiosperms or gymnosperms) (Aldinary et al., 2021).

Secondary metabolites are typically small organic compounds characterized by diverse chemical structures and biological functions, often possessing low molecular weight (Mosunova *et al.*, 2020). These compounds have had a profound impact on modern medicine, as evidenced by the fact that more than 52% of anti-infective small molecule drugs approved in recent decades originate from natural sources. Furthermore, approximately 42% of new drugs approved over the past 40 years are either natural products or their derivatives (Newman *et al.*, 2020). **Table 1**: Functional Group and Bioactive Compounds Produced by the Endophytic Fungi- Aspergillus fumigatus

Endophytic Fungus	Compound(s) Produced	Functional Group	Antimicrobial Activity	Reference
Aspergillus fumigatus	Linoleic acid, R-glycerol monolinoleate, bisdethio-(bis-methyl-thio)-gliotoxin, fumiquinazoline F, fumiquinazoline D, deoxythymidine, cerebroside A, (Z,Z)- N,N'-[1-[(4-hydroxyphenyl)methylene]- 2-[(4-methoxyphenyl)methylene]-1,2- ethanediyl]bisformamide, pyrazoline-3- one trimer, tricho-9-ene- 2α , 3α , 11α , 16 - tetraol	Alkaloids, Terpenoids, Steroids, Polyketones, Peptides, Flavonoids, Furandiones, Quinols, Perylene derivatives, Depsipeptides xanthones	Exhibits antimicrobial effects	Sharaff <i>et</i> <i>al.</i> , 2022
Aspergillus fumigatus	Hexadecanoic acid, Hexadecanoic acid methyl ester, bisabolol oxide B, 9,12- octadecadienoic acid, octadecenoic acid, 2-hydroxy-1-ethyl ester, linoelaidic acid, glycidyl palmitate, 9,17-octadecadienal, ethyl-9,12-octadecadienoate, glycidyl oleate, linoleoyl chloride	Alkaloids, Terpenoids, Steroids, Polyketones, Peptides, Flavonoids, Furandiones, Quinols, Perylene derivatives, Depsipeptides xanthones	Exhibits antimicrobial effects	Sharaff <i>et</i> <i>al.</i> , 2022

Sampling and Optimization of Media for the Production of Bioactive Molecules from Endophytic Fungi

In recent years, there has been considerable interest in the sampling and optimization of media for the production of bioactive compounds from endophytic fungi. These fungi are known to generate a wide range of bioactive compounds with diverse biological activities, including antibacterial, antifungal, antioxidant, anticancer, antidiabetic, and antiparasitic properties (Abdalla & McGraw, 2018). The potential of these bioactive compounds in the pharmaceutical industry are significant (Anand *et al.*, 2023).

Isolating endophytic fungi and bacteria with broad bioactivities requires careful consideration of

certain criteria, such as the ethnomedicinal history, habitat, unique biology, and endemism of the plant species. Additionally, it is generally recommended to select healthy-looking plants and young plant tissues over older materials (Yu et al., 2017). Various parts of plants, including seeds, leaves, stems, and bark, serve as reservoirs for endophytes. The process of isolating endophytes involves the chopping of plant material and surface sterilization. Surface sterilization is a critical step to eliminate external materials, molds, and contaminants, thereby promoting the growth of internal microorganisms. This is typically achieved by treating plant materials with surfactants such as ethanol (70-95%) and sodium hypochlorite (NaOCl 2-10%), followed by rinsing with ethanol

and sterile water to remove residual sterilants (Anand *et al.*, 2018).

To enhance the production of bioactive molecules, endophytic fungi can be cultured in optimized media (Table 2). The optimization of media composition, including factors such as carbon and nitrogen sources, pH, temperature, and other variables, is essential for maximizing the yield of

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bioactive compounds. Elicitors—biomolecules produced by endophytic fungi, such as polysaccharides, polypeptides, unsaturated fatty acids, and glycoproteins—play a role in protecting plants from pathogenic diseases and also promote the development of various phytochemicals, including alkaloids, flavonoids, terpenoids, saponins, and phenols (Singh & Kumar, 2023).

Table 2: Optimization of Culture Media and Conditions for Enhanced Production of Bioactive Compounds

 by Endophytic Fungi

Endophytic Fungi	Optimized Media/Conditions	Bioactive Compounds Produced	Key Findings	References
Aspergillus eucalypticola SBU-11AE	Potato Dextrose Agar (PDA), Potato Carrot Broth (PCB), Yeast Extract Peptone Dextrose (YEPD), Malt + Peptone; pH 5- 7		PCB media yielded the highest level of glabaridin $(7.26 \pm 0.44 \text{ mg/L})$, particularly at pH 6 due to starch and vitamin content supporting fungal growth and bioactive production	Parisa <i>et al.,</i> 2023
<i>Diaporthe</i> sp. GNBP-10	Potato Starch and Dextrose	Bis-anthraquinones	Optimized media with potato starch and dextrose produced the highest Bis-anthraquinones; minerals and excessive proteins had insignificant effects	Listiana <i>et al.</i> , 2023

The optimization of media composition, including carbon and nitrogen sources, pH, temperature, and other factors, plays a crucial role in maximizing the yield of bioactive molecules (Anand *et al.*, 2023).

Diverse Medicinal Potentials

The emergence of resistance among pathogenic microorganisms, the appearance of novel life-threatening viruses, and the increasing prevalence of communicable and non-communicable diseases present an urgent need to explore and utilize new resources that can provide sustainable solutions (Digra, 2023).

Historically, plants have served as a primary source of phytochemicals for the discovery and development of drugs (Adeleke & Babalola, 2021). Endophytes, microorganisms that live symbiotically within plants, and their bioactive byproducts are promising natural sources for future therapeutics. There has been growing interest in the medicinal potential of endophytes and their metabolites as biotherapeutic agents (Xia *et al.*, 2022).

Endophytic fungi are particularly remarkable for their ability to synthesize a diverse array of compounds with significant medicinal potential, including those with antiviral, antifungal, antibacterial, and anticancer properties. Many endophytic fungi are also potential sources of hormones and plant growth factors (Morales-Vargas, 2024). Additionally, some endophytes secrete extracellular enzymes, such as phosphatases, which convert insoluble phosphates into soluble forms, making them more accessible for plant uptake. The secondary metabolites produced by endophytic fungi enhance the host plant's immune system, which has been shown to reduce the severity of infections and mitigate damage caused by harmful microorganisms (Sharma *et al.*, 2021). The medicinal roles of endophytic fungi are detailed as follows:

A. Anticancer Compounds from Endophytic Fungi

According to the American Cancer Society (ACS), cancer ranks as the second leading cause of death worldwide. In 2020, the World Health Organization (WHO) reported that approximately 15 million people succumbed to malignant cells, and this number continues to rise. The most common cancers globally include breast, cervical, colon, prostate, oral, rectal, skin, and stomach cancers. The incidence of cancer varies significantly by region and country, with higher rates observed in more developed nations (Ali, 2024). Research is focused on developing innovative anticancer substances and treatments that specifically target cancer cells while minimizing side effects (Zayden et al., 2020). Various factors, such as genetics, lifestyle choices, and access to healthcare, influence the incidence of cancer (Iqbal et al., 2017). Table 3 present anticancer activity of secondary metabolites from Endophytic Fungi.

B. Antidiabetic Compounds from Endophytic Fungi

Diabetes is a metabolic disorder characterized by elevated blood glucose levels, commonly referred

to as blood sugar levels (Usman, 2024). Over time, this condition can lead to severe complications, including damage to the heart, blood vessels, eyes, kidneys, and nerves. Diabetes mellitus is particularly associated with various complications such as retinopathy, nephropathy, cardiovascular diseases, and neuropathy (Munandar, 2024). Secondary metabolites from Endophytic Fungi with Anti-Diabetic and Hypolipidemic Properties are presented in Table 4.

C. Antiviral Compounds from Endophytic Fungi

The continuous emergence of new virus serotypes, characterized by high mutation rates and low replication fidelity, complicates efforts to control these pathogens. With new viruses appearing daily, the current arsenal of vaccines and antivirals is becoming increasingly insufficient. The global challenge of drug resistance underscores the critical need for the development of novel antiviral compounds. Additionally, the growing resistance of human pathogens to existing antibiotics further highlights the urgency for alternative therapeutic strategies. Compounds derived from endophytic fungi present a vast, largely untapped reservoir of chemical structures with significant potential as sources of novel antiviral agents (Rashed, 2024) (Table 5). One of the major obstacles in developing effective antiviral drugs is the inadequacy, inappropriateness, or absence of robust antiviral screening methods.

	Endophytic Fungus	Host Plant	Compound(s) Produced	Cancer Type/Cell Line	Activity	Reference
393	Aspergillus terrus, Penicillium commune, P. chrysogenum, Talaromyces piophilus	Taxus brevifolia, Chilli plant	Taxol	MCF-7, HLK-210, HL-251	Cytotoxic activity	Kumar <i>et al.</i> , 2021
	Sinopodophyllum hexandrum, Dysosma veitchii	-	Podophyllotoxin	Leukemia, testicular, prostate, lung, ovarian	Effective in treating multiple cancers	Leung & Cassimeris, 2019
	Aspergillus niger	Camptotheca acuminata	Camptothecin	Colon cancer cell lines	Causes apoptosis, cell viability 11.85- 65.13%	Aswani & Soundhari, 2018
	Fusarium oxysporum, Alternaria sp.	Catharanthus roseus	Vinblastine	HepG-2	Effective at 7.48 μg/mL	Kousar <i>et al.</i> , 2022
	Pestalotiopsis microspora	Torreya taxifolia	Torreyanic acid	-	-	Deshmukh <i>et al.</i> , 2019
	Mucor fragilis	-	Kaempferol, podophyllotoxin	-	-	Deshmukh et al., 2019
	Xylaria sp. ZJWCF255	-	Cytochalasin Q	SMMC-772, MCF- 7, MGC80-3	Cytotoxic activity	Deshmukh et al., 2019
	Aspergillus terreus, A. flavus	Ficus elastica	-	MCF7, LS174 T, HCT29, HEPG-2	Possible anticancer properties	El-Sayed <i>et al.</i> , 2021
	Chaetomium globosum	-	Flavipin	HT-29, A549	Effective anticancer activity	Senthil <i>et al.</i> , 2019

Table 3: Anticancer Activity of Secondary Metabolites from Endophytic Fungi

Table 4: Secondary Metabolites from Endophytic Fungi with Anti-Diabetic and Hypolipidemic Properties

Endophytic Fungus	Host Plant	Compound(s) Produced	Biological Activity	Reference
Nigrospora oryzae	Combretum dolichopetalum	-	Lowering fasting blood sugar levels in diabetic mice	Uzor <i>et al.</i> , 2017
<i>Xylariaceae</i> spp.	<i>Quercus gilva</i> Blume	8-hydroxy-6,7-dimethoxy-3-methyl isocoumarins, abscisic acid, 70-hydroxy abscisic acid, 4-des-hydroxyl altersolanol A	Significant glucosidase inhibitory activity	Indrianingsih & Tachibana, 2017
<i>Fusarium</i> spp., <i>Alternaria</i> spp.	Rauwolfia densiflora, Leucas ciliata	-	Anti-diabetic properties	Adeleke <i>et al.</i> , 2021
Alternaria spp.	-	Glycoprotein (N-acetylgalactosamine)	In vivo and in vitro antidiabetic activity	-
Phoma spp., Aspergillus spp.	Salvadora oleoides	-	Anti-diabetic and hypolipidemic effect in Wistar albino rats	Ezekwesili & Ogbunugafor, 2015
Aspergillus sp.	Sonneratia apetala	Aspergiamides A and F	$\alpha\mbox{-glucosidase}$ inhibitory action with IC50 values of 40 and 83 μM	Ye et al., 2021
Aspergillus egypticus HT166S	Helianthus tuberosus	Polymethoxylated flavone (K-10)	Inhibition of α-amylase	Ruzieva <i>et al.</i> , 2022
Zasmidium sp. strain EM5-10	Mangrove	Tripalmitin	Significant inhibition of α- glucosidase activity	Lopez et al., 2019

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Table 5: Antiviral	Secondary	Metabolites	from	Endophytic Fungi

	Endophytic Fungus	Host Plant	Compound(s) Produced	Target Virus(es)	Biological Activity	Reference
4894	Cytonaema spp.	-	Cytonic acid A, Cytonic acid B	Human cytomegalovirus (HCMV)	Protease inhibition	Gupta <i>et al.</i> , 2020
	Trichoderma harzianum	Kadsura angustifolia	Nigranoic acid	HIV-1	Inhibits reverse transcriptase	Wang <i>et al.</i> , 2019
	Pestalotiopsis theae	-	Pestalotheol-C	HIV	Anti-HIV activities	Wang <i>et al.</i> , 2019
	Curvularia papendorfii	Vernonia amygdalina	Crude extract	Coronavirus	Reduces cytopathogenic effects by 40% at low MOI	Khiralla <i>et al</i> ., 2020
	Phoma sp.	Aconitum vilmorinianum	Phomanolide	-	-	Liu et al., 2019
	Jishengella endophytica	-	2-(furan-2-yl)-6-(2S, 3S, 4- trihydroxybutyl) pyrazine	Influenza A (H1N1)	Inhibits viral spread	Fadiji & Babalola, 2020
	<i>Alternaria</i> sp. QUE1Se	-	Altertoxins	HIV-1	Strong anti-HIV-1 effects	Raekiansyah <i>et</i> al., 2017
	<i>Emericella</i> spp. (HKZJ)	-	Emerimidine (A, B), Dehydroaustin, Austinol, Aspernidine (A, B), Austin, Emeriphenolicins (A, D), Acetoxydehydroaustin	Influenza A (H1N1)	Antiviral activity against influenza A virus	Raekiansyah <i>et al.</i> , 2017
	Alternaria alternata	-	Alternariol, Alternariol 9 Methyl Ether	Hepatitis C virus (HCV)	Inhibits HCV NS3/4a protease	El-Kassem <i>et</i> <i>al.</i> , 2019
	Cytonaema spp.	-	Cytonic acid A, Cytonic acid B	Human cytomegalovirus (HCMV)	Protease inhibition	Gupta <i>et al.</i> , 2020
	Trichoderma harzianum	Kadsura angustifolia	Nigranoic acid	HIV-1	Inhibits reverse transcriptase	Wang <i>et al.</i> , 2019
	Pestalotiopsis theae	-	Pestalotheol-C	HIV	Anti-HIV activities	Wang <i>et al.</i> , 2019
	Curvularia papendorfii	Vernonia amygdalina	Crude extract	Coronavirus	Reduces cytopathogenic effects by 40% at low MOI	Khiralla <i>et al.</i> , 2020
	Phoma sp.	Aconitum vilmorinianum	Phomanolide	-	-	Liu et al., 2019
	Jishengella endophytica	-	2-(furan-2-yl)-6-(2S, 3S, 4- trihydroxybutyl) pyrazine	Influenza A (H1N1)	Inhibits viral spread	Fadiji & Babalola, 2020
	<i>Alternaria</i> sp. QUE1Se	-	Altertoxins	HIV-1	Strong anti-HIV-1 effects	Raekiansyah <i>et</i> <i>al.</i> , 2017
	<i>Emericella</i> spp. (HKZJ)	-	Emerimidine (A, B), Dehydroaustin, Austinol, Aspernidine (A, B), Austin, Emeriphenolicins (A, D), Acetoxydehydroaustin	Influenza A (H1N1)	Antiviral activity against influenza A virus	Raekiansyah <i>et al.</i> , 2017
	Alternaria alternata	-	Alternariol, Alternariol 9 Methyl Ether	Hepatitis C virus (HCV)	Inhibits HCV NS3/4a protease	El-Kassem <i>et</i> <i>al.</i> , 2019

D. Antimicrobial Compounds from Endophytic Fungi

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The growing issue of antibiotic resistance among pathogenic bacteria necessitates an urgent search for new antimicrobial agents that are both effective and have fewer or milder side effects. Endophytic fungi, which can reside within plant tissues without causing any apparent harm to their hosts (Sharaf *et al.*, 2022), have emerged as promising sources of such agents (Table 6). These fungi are capable of synthesizing a wide range of secondary metabolites, including alkaloids, terpenoids, flavonoids, and polyketides, which hold significant potential as antimicrobial compounds (Awad, 2024).

E. Immunosuppressive Compounds from Endophytic Fungi

Immunosuppressive drugs are essential for preventing, suppressing, or minimizing organ rejection in transplant patients. They also play a crucial role in effectively managing autoimmune conditions such as lupus, psoriasis, insulindependent diabetes, and rheumatoid arthritis (Renda et al., 2021). Despite their importance, these drugs are often associated with significant side effects, underscoring the need for safer alternatives that can provide effective immune modulation with adverse fewer effects (Mohammedsaleh, 2022). Table 7 present a summary of immunosuppressive activity of Secondary Metabolites from Endophytic Fungi.

F. Antiparasitic Compounds from Endophytic Fungi

Protozoan parasites, such as *Trypanosoma cruzi*, *Plasmodium berghei*, *Plasmodium falciparum*, and *Leishmania amazonensis*, are responsible for a range of diseases, including Chagas disease, malaria, and leishmaniasis (Joselín *et al.*, 2021). These vector-borne illnesses are transmitted to humans through the bites or feeding processes of infected mosquitoes or flies. Predominantly affecting low-income countries, these diseases are classified by the WHO as neglected tropical diseases, receiving limited attention in terms of research and development. The treatments for these conditions face significant challenges, including low efficacy, toxicity, drug resistance, and high cost. Consequently, there is a pressing need to discover new drugs that are effective, safe, and affordable. To address these challenges, efforts are being directed toward various approaches, such as repurposing existing drugs, screening chemical libraries, and developing new candidates using targeted or natural product-based strategies (Table 8) (Capela *et al.*, 2019).

G. Antioxidant Compounds from Endophytic Fungi

Free radicals are unstable molecules generated either naturally through metabolic processes in the body or from external sources such as UV light, pesticides, medications, smoking, and alcohol (Dahan, 2023). These free radicals can damage cells, leading to various diseases, including cardiovascular disorders, diabetes, Down's syndrome, degenerative diseases, Alzheimer's disease, and Parkinson's disease (Saxena et al., 2021). Antioxidant compounds protect cells from damage by neutralizing free radicals, preventing them from causing cellular harm. These antioxidants are also crucial in cancer prevention, as they can interact with and neutralize the free radicals that contribute to the formation of cancer cells (Zhu et al., 2023).

The development of new antioxidant compounds is vital for improving human health and preventing degenerative and other diseases by stabilizing free radicals and reducing cellular damage. The demand for natural antioxidants has surged in recent years due to the growing market for nutritional supplements free from synthetic antioxidants (Gulchin, 2020). While antioxidants are found in various medicinal plants, vegetables, and fruits, the metabolites of fungal endophytes 4896

have also been identified as promising sources of new natural antioxidants (Sharaf *et al.*, 2022). Studies have shown that endophytic fungi can produce a wide range of substances with potent antioxidant properties (Table 9). Some of these Nig. J. Pure & Appl. Sci. Vol. 37 (Issue 1, 2024)

compounds, including phenolic acids, xanthones, flavonoids, terpenoids, and polyketides, have been successfully isolated and characterized (Mishra *et al.*, 2020).

Table 6: Antimicrobial	Secondary Metabolites	from Endophytic Fungi
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Endophytic Fungus	Host Plant	Compound(s) Produced	Target Pathogens	Biological Activity	Reference
<i>Aspergillus</i> sp. ASCLA	Callistemon subulatus	Isoshamixanthone	Staphylococcus aureus, Pseudomonas aeruginosa, Klebsiella pneumonia, etc.	Antibacterial activity	Kamel <i>et al.</i> , 2020
Aspergillus flavus	Ocimum basilicum	-	Staphylococcus aureus, Bacillus cereus, Bacillus subtilis, Escherichia coli, Salmonella typhimurium, Candida albicans, etc.	Antibacterial and antifungal properties	Sharaf <i>et al.</i> , 2022
Aspergillus nidulans	Ocimum basilicum	-	Various resistant microbes	Antibacterial properties	Elkhouly et al., 2021
Aspergillus fumigatus	Albizia lucidior	-	Escherichia coli, Staphylococcus aureus, Micrococcus luteus, Pseudomonas aeruginosa, Candida albicans	Antibacterial properties	Hussein <i>et</i> <i>al.</i> , 2022
Aspergillus niger, Emericella sp.	-	Cristatumin B, quiannulatic acid, dihydroauroglaucin	Multidrug-resistant Staphylococcus aureus (MDRSA), Enterococcus faecalis, Klebsiella pneumonia, Pseudomonas aeruginosa	Broad-spectrum antibacterial activity	Elkady <i>et al.</i> , 2022
Athelia rolfsii	Coleus amboinicus	Aromatic molecule with methoxy, hydroxyl, and methyl groups	Bacillus subtilis, Escherichia coli, Pseudomonas aeruginosa, Staphylococcus aureus, Streptococcus mutans	Potent antibacterial action	Astuti <i>et al.</i> , 2020
Aspergillus fumigatus	Ceriops decandra	Fumigatin, fraxetin	Escherichia coli, Micrococcus luteus, Staphylococcus aureus, Pseudomonas aeruginosa	Strong antibacterial activity	Zihad <i>et al.</i> , 2022
Alternaria spp.	Artemisia annua	Altersetin, n-butanol, ethylacetate	Various pathogenic gram- positive bacteria, phytopathogenic organisms	Strong antibacterial action, growth inhibition of phytopathogens	Fadiji & Babalola, 2020
Aspergillus tubenginsis	-	Anofinic acid	Gram-positive bacteria, gram- negative bacteria, unicellular fungi	Antibacterial activity, biofilm prevention	Mohamed <i>et</i> <i>al.</i> , 2020
Aspergillus versicolor	Pulicaria crispa (Forssk)	Aspergillethers A and B	Staphylococcus aureus, Bacillus cereus, Escherichia coli, Candida albicans, Geotrichium candidum	High antibacterial action	Maliehe <i>et</i> <i>al.</i> , 2022
Penicillium sp.	Stephania dielsiana	-	Various pathogenic bacteria	Potential antibacterial activity	Wu <i>et al</i> ., 2020
Penicillium citrinum	Azadirachta indica	-	Human pathogenic bacteria and fungi	Potential action	Kumari <i>et</i> <i>al.</i> , 2021

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Endophytic Fungus	Host Plant	Compound(s) Produced	Target Pathogens	Biological Activity	Reference
Penicillium ochrochloron	Taxus medium	-Pyrone derivatives	Various pathogenic bacterial and fungal strains	Antibacterial activity (MICs 12.5-100 µg/ml)	Zhao <i>et al.</i> , 2019
Aspergillus alternata	-	Ethyle acetate crude extract	Escherichia coli ATCC 11229, Bacillus subtilis RCMB 015, Staphylococcus aureus ATCC 25923, Candida albicans ATCC 90028	Promising antimicrobial activity	-
Aspergillus destruens	Calotropis gigantea	-	Wide variety of pathogenic bacteria strains	Suppressive activity	Elghaffar <i>et</i> <i>al.</i> , 2022
Fusarium spp.	-	-	Various bacterial and fungal strains	Antibacterial action	Elghaffar <i>et</i> <i>al.</i> , 2022
Trichoderma harzianum	Rosmarinus officinalis	-	Pseudomonas aeruginosa, Staphylococcus aureus, Klebsiella pneumonia, Bacillus subtilis, Escherichia coli	Growth inhibition	Abdulhadi <i>et</i> al., 2020
Trichoderma virens	Artemisia argyi	Trichocadinins B–G, cadinane-type sesquiterpene derivatives	-	Antibacterial properties	Shi <i>et al</i> ., 2019
Chaetomium globosum	-	Equisetin	Drug-resistant microorganisms	Antibacterial effectiveness	Zhao <i>et al.</i> , 2019
Xylaria ellisii	-	Ellisiiamide A	Escherichia coli	Bacterial activity	Ibrahim <i>et</i> <i>al.</i> , 2020
Diaporthe terebinthifolii	Schinus terebinthifolius	Orthosporin, diaporthin	-	Antibacterial properties	de Medeiros et al., 2018

Table 7: Immunosuppressive Activity of Secondary Metabolites from Endophytic Fungi

Endophytic Fungus	Host Plant	Compound(s) Produced	Immunosuppressive Activity	IC50/Effect	Reference
PGS1, NLL3	Psidium guajava, Newbouldia laevis	Citrinin, nidulalin, p- hydroxybenzoic acid, cyclopenin	Linked to immunosuppressive effects	-	Ujam <i>et al.</i> , 2021
Mycosphaerella nawae ZJLQ129	-	Mycousnine enamine	Specifically decreases T-cell proliferation	Blocks expression of CD25 and CD69	Wang <i>et al.</i> , 2017
Penicillium sp. ZJ- SY2	Sonneratia apetala	Peniphenone, methyl peniphenone, seven xanthones	Strong immunosuppressive effects	IC50: 5.9 to 9.3 µg/mL	Liu <i>et al.</i> , 2016

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Endophytic Fungus	Host Plant	Compound(s) Produced	Plasmodium Strain	IC50 Value (µg/mL)	Reference
Aspergillus terrus	Artemisia annua	-	Plasmodium falciparum	35.0	Kumarihamy <i>et al.</i> , 2019
Penicillium commune	Artemisia annua	-	Plasmodium falciparum	1.1	Kumarihamy <i>et al.</i> , 2019
Penicillium chrysogenum	Artemisia annua	-	Plasmodium falciparum	3.3	Kumarihamy <i>et al.</i> , 2019
Talaromyces piophilus	Artemisia annua	-	Plasmodium falciparum	7.6–9.9	Kumarihamy <i>et al.</i> , 2019
Nemania sp. UM10M	-	Epoxycytochalasin C	Plasmodium falciparum	Relatively weak	Kumarihamy <i>et al.</i> , 2019
Endophytic fungal strains IP-2 & IP-6	Artemisia annua	-	Plasmodium falciparum	30.0, 42.0	Kumarihamy <i>et al.</i> , 2019
Penicillium citrinum AMrb11	-	-	Plasmodium falciparum (Pf3D7 & PfINDO/PfDd2)	0.84–0.93	Ujam <i>et al.</i> , 2022
Neocosmospora rubicola AMb22	-	-	Plasmodium falciparum (Pf3D7 & PfINDO/PfDd2)	0.39–1.92	Ujam <i>et al.</i> , 2022
Neocosmospora rubicola	Azadirachta indica	-	Plasmodium falciparum	Dose- dependent	Ujam <i>et al</i> ., 2022
Aspergillus versicolor	-	-	Plasmodium falciparum	-	Toghueo <i>et al.</i> , 2019
Trichoderma afroharzianum	-	-	Plasmodium falciparum	-	Toghueo <i>et al.</i> , 2019
Fusarium sp.	-	-	Plasmodium falciparum	-	Toghueo <i>et al.</i> , 2019

Table 8: Antiplasmodial Activity of Secondary Metabolites from Endophytic Fungi

H. Antifungal Compounds from Endophytic Fungi

Pathogenic fungi are responsible for millions of deaths annually, as well as significant damage to agricultural products. The increasing resistance of these fungi to existing antifungal medications further exacerbates the issue (Almeida *et al.*,

2019). This growing problem underscores the urgent need for new bioactive compounds that are effective against resistant fungal strains. Among the promising candidates are bioactive compounds produced by endophytic fungi, including triterpenoids, sesquiterpenes, and diterpenoids, which have shown potential in combating plant fungal infections (Xu *et al.*, 2021).

Endo	phytic Fungus	Host Plant	Compound(s) Produced	Biological Activity	Reference
Epicoccum nigrum	ccum nigrum	Acanthus ilicifolius L.	Bioactive substances	Antioxidant and α- glucosidase inhibitory action	Yan <i>et al.</i> , 2020
Asper	gillus oryzae DK7	<i>Mangifera casturi</i> Kosterm	-	Antioxidant activity (IC50: 142.96 μg/mL)	da Silva <i>et</i> <i>al.</i> , 2020
Asper AKF1	gillus minisclerotigens	<i>Mangifera casturi</i> Kosterm	-	Antioxidant activity (IC50: 145.01 μg/mL)	da Silva <i>et</i> <i>al.</i> , 2020
Penici	<i>illium</i> spp.	-	Ferulic acid, cinnamic acid, quercetin, rutin	Antioxidant activity	Nischitha <i>et</i> <i>al.</i> , 2021
Curvu	<i>llaria</i> sp.	-	Antioxidants	Antioxidant activity	Kalimuthu e al., 2022
Curvu	laria lunata	-	Antioxidants	Antioxidant activity	Ye <i>et al</i> ., 2021
Collet	<i>otrichum</i> sp.	-	Antioxidants	Antioxidant activity	Palupi <i>et al</i> . 2021
Clado	sporium tenuissimum	-	Antioxidants	Antioxidant activity	Fadhillah <i>et</i> <i>al</i> ., 2019
Vario	us endophytic fungi	Medicinal plants from Central Sulawesi	-	High antioxidant and antibacterial activities	Raunsai <i>et</i> <i>al.</i> , 2018
-	11, LO 261, FE 082,	Albizia zygia, Millettia thonningii, Alchornea cordifolia, Ficus exasperata	Pyrogallol, dl-alpha- tocopherol, Alpha tocospiro, Linoleic acid, 9- octadecenamide, lupeol, 9- octadecenoic acid (Z)	Antioxidant activity	Ibrahim <i>et</i> <i>al.</i> , 2021
Altern	aria alternata	Azadirachta indica	-	Antioxidant activity (IC50: 38 μg/mL)	Chatterjee <i>et</i> <i>al.</i> , 2019
Penici	illium citrinum	Azadirachta indica	-	Antioxidant activity (IC50: 52.13 μg/mL)	Chatterjee <i>el</i> <i>al.</i> , 2019
Collet	omium sp., otrichum sp., daria sp., Trichoderma	Related host plants	Antioxidants	Antioxidant activity (31- 69%)	Kumaresan <i>et al.</i> , 2015
<i>Cui ru</i>					

Table 9: Antioxidant Secondary Metabolites from Endophytic Fungi

	Endophytic Fungus	Host Plant	Compound(s) Produced	Target Fungus	Biological Activity	Reference
00	Xylaria sp.	Paullinia cupana (Guarana plant)	Cytochalasin D	Colletotrichum gloeosporioides	Fungistatic action	Elias <i>et al.</i> , 2018
	<i>Mucor</i> sp.	Centaurea stoebe	14- hydroxyterezine D, Terezine E	Aspergillus terreus	Weak antifungal activity (MICs: 127.8 µg/ml and 111.2 µg/ml, respectively)	Abdou <i>et</i> <i>al.</i> , 2018
	Trichoderma koningiopsis YIM PH 30002	Panax notoginseng	Koningiopisin C	Plectosphaerella cucumerina	Antifungal activity (MIC: 57.1 μg/ml)	Liu <i>et al.</i> , 2016
	<i>Drechslera</i> sp. strain 678	Neurachne alopecuroidea (Australian grass)	-	Various plant pathogens	Broad-spectrum potency against plant diseases	d'Errico <i>et</i> <i>al.</i> , 2020
	Colletotrichum gloeosporioides	Artemisia mongolica	Colletotric acid	Various fungi	Antifungal properties	Li <i>et al.</i> , 2019
	-	-	Roridoxins A and C	Alternaria tenuissima, A. niger, Pyricularia grisea, Fusarium oxysporum	Significant antifungal effects	Li <i>et al.</i> , 2019
	-	-	Trichobreols	Pathogenic yeast-like fungi	Effective antifungal efficacy	Yamazaki et al., 2020
	Aspergillus spp., Penicillium spp., Trichoderma viride	-	Synthetic griseofulvin derivatives	Various fungi	More effective than bifonazole and ketoconazole (3–16 fold and 7–42 times, respectively)	Kartsev et al., 2019
	<i>Aspergillus</i> sp.	Brassica napus (Oilseed rape plant)	Penicillither, Methyl dichloroasterrate	Botrytis cinerea, Monilinia fructicola, Sclerotinia sclerotiorum, S. trifoliorum	Inhibitory effects (EC50s: 21.7 to 151.2 µg/ml)	Qin <i>et al.</i> , 2019
	<i>Phomopsis</i> sp. YE3250	<i>Paeonia delavayi</i> (Medicinal plant)	Phomopoxides B, C, D, F, G	Candida albicans, Aspergillus niger, Pyricularia oryzae, Fusarium avenaceum, Hormodendrum compactum	Antifungal activity (MICs: 46.5-372.1 μg/ml)	Huang <i>et</i> <i>al.</i> , 2018

Table 10: Antifungal Secondary Metabolites from Endophytic Fungi

Conclusion

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Endophytic fungi represent a promising and largely untapped source of bioactive compounds with significant therapeutic potential. This review has highlighted the broad spectrum of medicinal compounds produced by endophytic fungi, including anticancer, antidiabetic, antiviral, antimicrobial, immunosuppressive, antiparasitic, antioxidant, and antifungal agents. The ability of these fungi to synthesize novel and potent bioactive molecules positions them as invaluable resources in the discovery and development of new therapeutic agents.

Despite the promising in vitro activities demonstrated by these compounds, transitioning from laboratory research to clinical applications remains a substantial challenge. Future research should focus on optimizing extraction and purification processes, elucidating mechanisms of action, and conducting comprehensive in vivo studies to establish the safety and efficacy of these compounds. Additionally, advancements in biotechnology and synthetic biology could be leveraged to enhance the production and scalability of these bioactive compounds, potentially accelerating their integration into clinical practice.

Endophytic fungi hold great promise as sustainable sources of new pharmaceuticals. Continued exploration and development of these fungi could lead to innovative solutions to some of the most pressing health challenges, potentially revolutionizing future therapeutic practices.

References

- Abdalla, M., & McGaw, L. (2018). Bioprospecting of South African plants as a unique resource for bioactive endophytic microbes. *Frontiers in Pharmacology*, 9. https://doi.org/10.3389/fphar.2018.00456
- Abdelaziz, A. M., El-Wakil, D. A., Attia, M. S., Ali, O. M., AbdElgawad, H., & Hashem, A. H. (2022).
 Inhibition of *Aspergillus flavus* growth and aflatoxin production in *Zea mays* L. using

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endophytic Aspergillus fumigatus. Journal of Fungi, 8(5), 482.

- Abdou, R., Shabana, S., & Rateb, M. E. (2018). Terezine E, bioactive prenylated tryptophan analogue from an endophyte of *Centaurea* stoebe. Natural Product Research, 34, 503–510. https://doi.org/10.1080/14786419.2018.1489393
- Abdulhadi, S. Y., Hasan, G. Q., & Gergees, R. N. (2022). Molecular detection and antimicrobial activity of endophytic fungi isolated from a medicinal plant *Rosmarinus officinalis*. *Annals of Tropical Medicine and Public Health*, 23, 231– 384.
- Adeleke, B. S., & Babalola, O. O. (2021). Pharmacological potential of fungal endophytes associated with medicinal plant: Review. *Journal* of Fungi, 7, 147.
- Adeleke, B. S., & Babalola, O. O. (2021). Plant growthpromoting root-colonizing bacterial endophytes. *Rhizosphere*, 20, 100433. <u>https://doi.org/10.1016/j.rhisph.2021.100433</u>
- Aisyah, N., Zulkifli, L., & Rasmi, D. (2023). Isolation of endophytic bacteria and fungi from soursop (Annona muricata L.) and bioactivity test as antimicrobial against Escherichia coli, Staphylococcus epidermidis, and Candida albicans. Jurnal Pijar Mipa, 18(2), 254-259. https://doi.org/10.29303/jpm.v18i2.3834
- Aldinary, A. M., Abdelaziz, A. M., Farrag, A. A., & Attia, M. S. (2021). Biocontrol of tomato Fusarium wilt disease by a new Moringa endophytic Aspergillus isolates. Materials Today: Proceedings. https://doi.org/10.1016/j.matpr.2021.03.423
- Ali, H., Ishtiaq, R., Tedder, B., Zweigle, J., Nomigolzar, R., Dahiya, D. S., ... & Tillmann, H. L. (2024). Trends in mortality from gastrointestinal, hepatic, and pancreatic cancers in the United States: A comprehensive analysis (1999–2020). *JGH Open, 8*(4). https://doi.org/10.1002/jgh3.13064
- Almeida, F., Rodrigues, M. L., & Coelho, C. (2019). The still underestimated problem of fungal

Nig. J. Pure & Appl. Sci. Vol. 37 (Issue 1): 4887-4909

diseases worldwide. Frontiers in Microbiology, 10, 214.

- Anand, K., Kumar, V., Prasher, I., Sethi, M., Raj, H., Ranjan, H., & Pandey, G. (2023). Bioactive molecules from fungal endophytes and their applications in pharmaceutical industries: Challenges and future scope. *Zeitschrift Für Allgemeine Mikrobiologie*, 63(7), 690-708. https://doi.org/10.1002/jobm.202200696
 - Anand, U., Pal, T., Yadav, N., Singh, V. K., Tripathi, V., & Choudhary, K. K. (2023). Current scenario and future prospects of endophytic microbes: Promising candidates for abiotic and biotic stress management for agricultural and environmental sustainability. *Microbial Ecology*. https://doi.org/10.1007/s00248-023-02190-1
 - Astuti, P., Rollando, R., Pratoko, D. K., & Wahyuono, S., & Nurrochmad, A. (2020). Antimicrobials and cytotoxic activities of a compound produced by an endophytic fungus isolated from the leaves of *Coleus amboinicus* Lou. *International Journal* of *Pharmaceutical Research*, 13, 2632–2644.
 - Aswani, A., & Soundhari, C. (2018). Production of camptothecin from endophytic fungi and characterization by high-performance liquid chromatography and anticancer activity against colon cancer cell line. Asian Journal of Pharmaceutical and Clinical Research, 11, 166. https://doi.org/10.22159/ajpcr.2018.v11i3.18921
 - Awad, L. (2024). First record of endophytic fungi *Thielaviabasicola* isolated from xerophyte plants *Suaeda aegyptiaca* in Basra, Iraq. *IOP Conference Series: Earth and Environmental Science,* 1371(3), 032016. <u>https://doi.org/10.1088/1755-</u> <u>1315/1371/3/032016</u>
 - Bamisile, B. S., Dash, C. K., Akutse, K. S., Keppanan, R., & Wang, L. D. (2018). Fungal endophytes: Beyond herbivore management. *Frontiers in Microbiology*, 9, 11. https://doi.org/10.3389/fmicb.2018.00544
 - Bernardini, S., Tiezzi, A., Laghezza Masci, V., & Ovidi, E. (2018). Natural products for human health: An historical overview of the drug discovery

approaches. *Natural Product Research, 32,* 1926–1950. https://doi.org/10.1080/14786419.2017.1356838

- Bezerra, J., Nascimento, C., Barbosa, R., Silva, D., Svedese, V., Silva-Nogueira, E., ... & Souza-Motta, C. (2015). Endophytic fungi from medicinal plant *Bauhinia forficata*: Diversity and biotechnological potential. *Brazilian Journal of Microbiology*, 46(1), 49-57. <u>https://doi.org/10.1590/s1517-</u> 838246120130657
- Bonaterra, A., Badosa, E., & Daranas, N. (2022). Bacteria as biological control agents of plant diseases. *Microorganisms*, 10, 1759. <u>https://doi.org/10.3390/microorganisms1009175</u> <u>9</u>
- Capela, R., Moreira, R., & Lopes, F. (2019). An overview of drug resistance in protozoal diseases. *International Journal of Molecular Sciences*, 20, 5748.
- Chatterjee, S., Ghosh, R., & Mandal, N. C. (2019). Production of bioactive compounds with bactericidal and antioxidant potential by endophytic fungus *Alternaria alternata* AE1 isolated from *Azadirachta indica* A. Juss. *PLOS ONE*, 14, e0214744.
- d'Errico, G., Aloj, V., Flematti, G. R., Sivasithamparam, K., Worth, C. M., & Lombardi, N. (2020). Metabolites of a Drechslera sp. endophyte with potential as biocontrol and bioremediation agent. Natural Product Research. Online ahead of print. https://doi.org/10.1080/14786419.2020.1737058
- da Silva, M. H. R., Cueva-Yesquén, L. G., & Júnior, S. B. (2020). Endophytic fungi from *Passiflora incarnata*: An antioxidant compound source. *Archives of Microbiology*, 202(10), 2779–2789. <u>https://doi.org/10.1007/s00203-020-02001-v</u>
- Dadwal, V. (2024). Recent progress in metabolomics research: Insights into fruit metabolome, harvesting techniques, and health-promoting bioactives. *International Journal of Food Science* & Technology, 59(8), 5285–5302. https://doi.org/10.1111/ijfs.17308

4902

- Dahan, S., Khalouf, R., Salcedo, N., Taieb, C., & Gassia, V. (2023). Knowledge of free radicals and antioxidants in dermatology in the general population. *JEADV Clinical Practice*, *3*(1), 142–149. https://doi.org/10.1002/jvc2.274
- de Medeiros, A. G., Savi, D. C., & Mitra, P. (2018). Bioprospecting of *Diaporthe terebinthifolii* LGMF907 for antimicrobial compounds. *Folia Microbiologica*, 63(4), 499–505. https://doi.org/10.1007/s12223-018-0587-2
- de Paula Nogueira Cruz, F., Bogas, A. C., & Paiva de Sousa, C. (2021). Plant-associated microorganisms as a potent bio-factory of active molecules against multiresistant pathogens. In *Antimicrobial Resistance—A One Health Perspective*. IntechOpen. https://doi.org/10.5772/intechopen.97493
- Deshmukh, S. K., Gupta, M. K., Prakash, V., & Reddy, M. S. (2019). Fungal endophytes: A novel source of cytotoxic compounds. In S. Jha (Ed.), *Endophytes and Secondary Metabolites* (pp. 365–426). Springer. https://doi.org/10.1007/978-3-030-11992-6 9
- Digra, S., & Nonzom, S. (2023). An insight into endophytic antimicrobial compounds: An updated analysis. *Plant Biotechnology Reports*. https://doi.org/10.1007/s11816-023-00752-5
- Eid, A. M., Fouda, A., & Abdel-Rahman, M. A. (2021). Harnessing bacterial endophytes for promotion of plant growth and biotechnological applications: An overview. *Plants*, *10*(5), 935. https://doi.org/10.3390/plants10050935
- Elghaffar, R. Y. A., Amin, B. H., Hashem, A. H., & Sehim, A. E. (2022). Promising endophytic *Alternaria alternata* from leaves of *Ziziphus spina-christi*: Phytochemical analyses, antimicrobial and antioxidant activities. *Applied Biochemistry and Biotechnology*, 194(9), 3984– 4001. https://doi.org/10.1007/s12010-021-03652-8
- Elias, L. M., Fortkamp, D., Sartori, S. B., Ferreira, M. C., Gomes, L. H., & Azevedo, J. L. (2018). The potential of compounds isolated from *Xylaria* spp. as antifungal agents against anthracnose.

Brazilian Journal of Microbiology, 49(1), 840–847. https://doi.org/10.1016/j.bjm.2018.03.003

- Elkady, W. M., Raafat, M. M., Abdel-Aziz, M. M., Al-Huqail, A. A., Ashour, M. L., & Fathallah, N. (2022). Endophytic fungus from *Opuntina fiscusindica*: A source of potential bioactive antimicrobial compounds against multidrugresistant bacteria. *Plants*, *11*(7), 1070. https://doi.org/10.3390/plants11081070
- El-Kassem, L. A., Hawas, U. W., El-Souda, S., Ahmed,
 E. F., El-Khateeb, W., & Fayad, W. (2019). Anti-HCV protease potential of endophytic fungi and cytotoxic activity. *Biocatalysis and Agricultural Biotechnology*, 19, 101170. https://doi.org/10.1016/j.bcab.2019.101170
- Elkhouly, H. I., Hamed, A. A., El Hosainy, A. M., Ghareeb, M. A., & Sidkey, N. M. (2021).
 Bioactive secondary metabolite from endophytic *Aspergillus tubenginses* ASH4 isolated from *Hyoscyamus muticus*: Antimicrobial, antibiofilm, antioxidant and anticancer activity. *Pharmacognosy Journal*, 13(2), 434. https://doi.org/10.5530/pj.2021.13.55
- El-Sayed, A. S. A., Khalaf, S. A., Azez, H. A., Hussein, H. A., El-Moslamy, S. H., Sitohy, B., & El-Baz, F. (2021). Production, bioprocess A. optimization and anticancer activity of Camptothecin from Aspergillus terreus and Aspergillus flavus, endophytes of Ficus elastica. Process Biochemistry, 107, 59-73. https://doi.org/10.1016/j.procbio.2021.05.007
- Fadhillah, F., Yohandini, H., & Widjajanti, H. (2019). Chemical compound isolated from antioxidant active extract of endophytic fungus *Cladosporium tenuissimum* in *Swietenia mahagoni* leaf stalks. *Biodiversitas Journal of Biological* <u>Diversity.</u> <u>https://doi.org/10.13057/biodiv/d200929</u>
- Fadiji, A. E., & Babalola, O. O. (2020). Elucidating mechanisms of endophytes used in plant protection and other bioactivities with multifunctional prospects. *Frontiers in Bioengineering and Biotechnology*, 8, 467. https://doi.org/10.3389/fbioe.2020.00467

- Gulchin, I. (2020). Antioxidants and antioxidant methods: An updated overview. Archives of Toxicology, 94(3), 651–715. https://doi.org/10.1007/s00204-019-02650-6
- 4904 Gupta, S., Chaturvedi, P., Kulkarni, M. G., & Van Staden, J. (2020). A critical review on exploiting the pharmaceutical potential of plant endophytic fungi. *Biotechnology Advances, 39*, 107462. https://doi.org/10.1016/j.biotechadv.2020.10746 2
 - Hastuti, U., Sulisetijono, S., Rahmawati, D., Sari, R., Hartono, S., Thoyibah, C., & Pratiwi, S. (2021).
 Identification and histological observation of endophytic fungi isolated from medicinal plant, *Physalis angulata* L. *Hayati Journal of Biosciences*, 28(2), 130. https://doi.org/10.4308/hjb.28.2.130
 - Huang, R., Jiang, B. G., Li, X. N., Wang, Y. T., Liu, S. S., & Zheng, K. X. (2018). Polyoxygenated cyclohexenoids with promising α-glycosidase inhibitory activity produced by *Phomopsis* sp. YE3250, an endophytic fungus derived from *Paeonia delavayi. Journal of Agricultural and Food Chemistry*, 66(5), 1140–1146. https://doi.org/10.1021/acs.jafc.7b04998
 - Hussaini, I., Ahmed, H., Ahmad, H., Sulaiman, M., & Usman, A. (2022). Preliminary screening for antibacterial activity of endophytic fungi isolated from Azadirachta indica and Mentha piperita against Staphylococcus aureus, Escherichia coli and Pseudomonas aeruginosa. Algerian Journal of Biosciences, 3(2), 056–060. https://doi.org/10.57056/ajb.v3i2.57
 - Hussein, M. E., Mohamed, O. G., El-Fishawy, A. M., El-Askary, H. I., El-Senousy, A. S., El-Beih, A. A., Nossier, E. S., Naglah, A. M., Almehizia, A.
 A., Tripathi, A., & Hamed, A. A. (2022). Identification of antibacterial metabolites from endophytic fungus *Aspergillus fumigatus*, isolated from *Albizia lucidior* leaves (Fabaceae), utilizing metabolomic and molecular docking techniques. *Molecules*, 27(3), 1117. https://doi.org/10.3390/molecules27031117
 - Ibrahim, A., Tanney, J. B., Fei, F., Seifert, K. A., Cutler, G. C., Capretta, A., & Sumarah, M. W. (2020).

Metabolomic-guided discovery of cyclic nonribosomal peptides from *Xylaria ellisii* sp. nov., a leaf and stem endophyte of *Vaccinium angustifolium*. *Scientific Reports*, *10*(1), 4599. https://doi.org/10.1038/s41598-020-61577-8

- Ibrahim, M., Oyebanji, E., Fowora, M., Aiyeolemi, A., Orabuchi, C., Akinnawo, B., & Adekunle, A. A. (2021). Extracts of endophytic fungi from leaves of selected Nigerian ethnomedicinal plants exhibited antioxidant activity. *BMC Complementary Medicine and Therapies*, 21(1), 98. <u>https://doi.org/10.1186/s12906-021-03269-3</u>
- Indrianingsih, A. W., & Tachibana, S. (2017). α-Glucosidase inhibitor produced by an endophytic fungus, *Xylariaceae* sp. QGS 01 from *Quercus* gilva Blume. Food Science and Human Wellness, 6(2), 88–95. https://doi.org/10.1016/j.fshw.2017.05.001
- Iqbal, J., Abbasi, B. A., Mahmood, T., Kanwal, S., Ali, B., & Khalil, A. T. (2017). Plant-derived anticancer agents: A green anticancer approach. *Asian Pacific Journal of Tropical Medicine*, 7(12), 1129–1150. https://doi.org/10.1016/S1995-7645(14)60200-1
- Jha P., Kaur Tamanna, Chhabra Ishita, Panja Avirup, Paul Sushreeta, Kumar Vijay, Malik Tabarak (2023). Endophytic fungi: hidden treasure chest of antimicrobial metabolites interrelationship of endophytes and metabolites. *Frontiers in Microbiology*, vol. 14.
- Joselín, D., Martínez, I., Rodríguez-Moreno, Á., Ghiaur, G., Isaias, L., Sánchez-Cordero, V., & Espinoza, B. (2021). Multiple discrete typing units of *Trypanosoma cruzi* infect sylvatic *Triatoma dimidiata* and *Panstrongylus rufotuberculatus* in southeast Mexico. *American Journal of Tropical Medicine and Hygiene*, 105(4), 1042-1049. https://doi.org/10.4269/ajtmh.20-1574
- Kalimuthu AK, Parasuraman P, Sivakumar P, Murugesan S, Arunachalam S, Pandian SRK, Ravishankar V, Ammunje DN, Sampath M, Panneerselvam T, Kunjiappan S. (2022). In silico, in vitro screening of antioxidant and anticancer potentials of bioactive secondary metabolites from an endophytic fungus

(*Curvularia* sp.) from *Phyllanthus niruri* L. *Environmental Science and Pollution Research*. https://doi.org/10.1007/s11356-022-19249-0.

- Kamel RA, Abdel-Razek AS, Hamed A, Ibrahim RR, Stammler HG, Frese M, Sewald N, Shaaban M. (2020). Isoshamixanthone: a new pyrano xanthone from endophytic *Aspergillus* sp. ASCLA and absolute configuration of epiisoshamixanthone. *Natural Product Research*; 34(8):1080–90.
 - Kartsev, V., Geronikaki, A., Petrou, A., Lichitsky, B., Kostic, M., Smiljkovic, M. (2019). Griseofulvin derivatives: synthesis, molecular docking and biological evaluation. *Current Topics in Medicinal Chemistry*, 19, 1145–1161. doi: 10.2174/1568026619666190523080136
 - Khiralla A, Spina R, Varbanov M, Philippot S, Lemiere P, Slezack-Deschaumes S, André P, Mohamed I, Yagi SM, Laurain-Mattar D. (2020). Evaluation of antiviral, antibacterial and antiproliferative activities of the endophytic fungus *Curvularia papendorfii*, and isolation of a new polyhydroxyacid. *Microorganisms*; 8(9):1353.
 - KouipouToghueo, R. M., & Boyom, F. F. (2019). Endophytic fungi from *Terminalia* species: a comprehensive review. *Journal of Fungi*, 5(2), 43.
 - Kumar S, Aharwal RP, Jain R, Sandhu SS. (2021). Bioactive molecules of endophytic fungi and their potential in anticancer drug development. *Current Pharmacology Reports*; 7(2):27–41.
 - Kumaresan, S.; Karthi, V.; Senthilkuma, V.; Balakumar, B.S.; Balakumar, A. (2015). Biochemical constituents and antioxidant potential of endophytic fungi isolated from the leaves of *Azadirachta indica* A. Juss (Neem) from Chennai, India. Journal of Academic and Industrial Research, 3, 355–361.
 - Kumari P, Singh A, Singh DK, Sharma VK, Kumar J, Gupta VK, Bhattacharya S, Kharwar RN. (2021).
 Isolation and purification of bioactive metabolites from an endophytic fungus *Penicillium citrinum* of *Azadirachta indica*.

South African Journal of Botany, 139:449–57. https://doi.org/10.1016/j.sajb.2021.02.020.

- Kumarihamy, M.; Ferreira, D.; Croom, E.C.M., Jr.; Sahu, R.; Tekwani, B.L.; Duke, S.O.; Khan, S.; Techen, N.; Nanayakkara, N.P.D. (2019). Antiplasmodial and cytotoxic cytochalasins from an endophytic fungus, *Nemania* sp. UM10M, isolated from a diseased *Torreya taxifolia* leaf. *Molecules*, 24, 777.
- Leung JC, Cassimeris L (2019). Reorganization of paclitaxel-stabilized microtubule arrays at mitotic entry: roles of depolymerizing kinesins and severing proteins. *Cancer Biology & Therapy*, 20:1337–1347. https://doi.org/10.1080/15384047.2019.1638678
- Li, T. X., Xiong, Y. M., Chen, X., Yang, Y. N., Wang, Y., Jia, X. W., (2019). Antifungal macrocyclic trichothecenes from the insect-associated fungus *Myrothecium roridum. Journal of Agricultural* and Food Chemistry, 67, 13033–13039. doi: 10.1021/acs.jafc.9b04507
- Lin H, Liang Y, Kaliaperumal K, Xiong Q, Duan S, Jiang Y, Zhang J. (2023). Linoleic acid from the endophytic fungus *Diaporthe* sp. HT-79 inhibits the growth of *Xanthomonas citri* subsp. *citri* by destructing the cell membrane and producing reactive oxygen species (ROS). *Pesticide Biochemistry and Physiology*. doi: 10.1016/j.pestbp.2023.105423. PMID: 37105613.
- Liu S-S, Jiang J-X, Huang R (2019). A new antiviral 14nordrimane sesquiterpenoid from an endophytic fungus *Phoma* sp. *Phytochemistry Letters*, 29:75–78. https://doi.org/10.1016/j.phytol.2018.11.005
- Liu, H.; Chen, S.; Liu, W.; Liu, Y.; Huang, X.; She, Z. (2016). Polyketides with immunosuppressive activities from mangrove endophytic fungus *Penicillium* sp. ZJ-SY2. *Marine Drugs*, 14, 217.
- Lopéz, D.; Cherigo, L.; Mejia, L.C.; Loza-Mejía, M.A.; Martínez-Luis, S. (2019). α-Glucosidase inhibitors from a mangrove-associated fungus, *Zasmidium* sp. strain EM5-10. *BMC Chemistry*, 13, 22.

4905

- Maliehe TS, Mbambo M, Nqotheni MI, Senzo NS, Shandu JSE (2022). Antibacterial effect and mode of action of secondary metabolites from fungal endophyte associated with *Aloe ferox* Mill. *Microbiology Research*, 13(1):90–101.
- Mashar, H., Supriyono, T., Ismail, I., Yunita, S., & Dali, D. (2023). Exploration of endophytic fungi in the kelakai (*Stenochlaena palustris*) and their antibacterial effect. *International Journal of Pharmaceutical Quality Assurance*, 14(02), 250-254. <u>https://doi.org/10.25258/ijpqa.14.2.01</u>
- Meshram, V.; Gupta, M. (2019). Endophytic fungi: A quintessential source of potential bioactive compounds. In Sustain: Endophytes for a Growing World; Hodkinson, T., Doohan, F., Saunders, M., Murphy, B., Eds.; Cambridge University Press: Cambridge, UK, Volume 13, pp. 277–309.
- Mishra, R.C.; Goel, M.; Barrow, C.J.; Deshmukh S.K. (2020). Endophytic fungi: An untapped source of potential antioxidants. *Current Bioactive Compounds*, 16, 944–964.
- Mohamed GA, Ibrahim SRM, Asfour HZ. (2020). Antimicrobial metabolites from the endophytic fungus Aspergillus versicolor. Phytochemistry Letters, 35:152–5. https://doi.org/10.1016/j.phytol.2019.12.003.
- Mohammedsaleh, Z.M. (2022). The use of patientspecific stem cells in different autoimmune diseases. *Saudi Journal of Biological Sciences*, 29, 3338–3346.
- Morales-Vargas, A. (2024). Endophytic fungi for crops adaptation to abiotic stresses. <u>https://doi.org/10.20944/preprints202402.1287.v</u> <u>1</u>
- Mosunova O., Navarro-Muñoz J.C., Collemare J. (2020). The biosynthesis of fungal secondary metabolites: From fundamentals to biotechnological applications. In *Reference Module in Life Sciences*. Elsevier; Amsterdam, The Netherlands.
- Munandar, I. (2024). The relationship between the duration of diabetes mellitus and diabetic neuropathy in the internal medicine polyclinic of

Linggajati General Hospital. *Blantika Multidisciplinary Journal*, 2(8). https://doi.org/10.57096/blantika.v2i8.179

- Nalin Rathnayake, G.R., Savitri Kumar, N., Jayasinghe, L. (2019). Secondary Metabolites Produced by an Endophytic Fungus *Pestalotiopsis microspora*. *Natural Products and Bioprospecting*, 9, 411– 417. <u>https://doi.org/10.1007/s13659-019-00225-</u>0
- Newman D.J., Cragg G.M. (2020). Natural Products as Sources of New Drugs over the Nearly Four Decades from 01/1981 to 09/2019. *Journal of Natural Products*, 83, 770–803. https://doi.org/10.1021/acs.jnatprod.9b01285
- Nischitha R, Shivanna MB. (2021). Metabolite Fingerprinting, In Vitro Antimicrobial and Antioxidant Activities and In-silico Docking in Alloteropsis cimicina and its Endophytic Fungus Penicillium pinophilum. Molecular Biology Reports, 48(5), 4021–4037. https://doi.org/10.1007/s11033-021-06410-0
- Noemi Carla Baron, Everlon Cid Rigobelo. (2022). Endophytic Fungi: A Tool for Plant Growth Promotion and Sustainable Agriculture. *Mycology*, 13(1), 39–55. https://doi.org/10.1080/21501203.2021.1945699
- Nurunnabi TR, Sabrin F, Sharif DI, Nahar L, Sohrab MH, Sarker SD, Rahman SMM, Billah MM. (2020). Antimicrobial Activity of Endophytic Fungi Isolated from the Mangrove Plant Sonneratia apetala (Buch.-Ham) from the Sundarbans Mangrove Forest. Advanced Traditional Medicine, 20(3), 419–425. https://doi.org/10.1007/s13596-019-00422-9
- Palupi KD, Ilyas M, Agusta A. (2021). Endophytic Fungi Inhabiting *Physalis angulata* L. Plant: Diversity, Antioxidant, and Antibacterial Activities of Their Ethyl Acetate Extracts. *Journal of Basic and Clinical Physiology and Pharmacology*, 32(4), 823–829.
- Pang, L. (2023). Biosyntheses of Azetidine-Containing Natural Products. Organic & Biomolecular Chemistry, 21(36), 7242–7254. https://doi.org/10.1039/d3ob01205k

- Parisa B. G., Mohsen F., Mohammad H. M. (2023). Development and Optimization of Culture Medium for the Production of Glabridin by *Aspergillus eucalypticola*: An Endophytic Fungus Isolated from *Glycyrrhiza glabra* L. (Fabaceae). *Mycobiology*. https://doi.org/10.1080/12298093.2023.2225253
- Pasrija, P., Girdhar, M., Kumar, M., Arora, S., Katyal, A. (2022). Endophytes: An Unexplored Treasure to Combat Multidrug Resistance. *Phytomedicine Plus*, 2, 100249.
- Qin, J., Lyu, A., Zhang, Q.H., Yang, L., Zhang, J., Wu, M.D. (2019). Strain Identification and Metabolites Isolation of Aspergillus capensis CanS-34A from Brassica napus. Molecular Biology Reports, 46, 3451–3460. https://doi.org/10.1007/s11033-019-04808-5
- Rajakumari, V., Rajagopal, K., Mohanasundaram, S. (2018). Various Chemical Groups Produced by Endophytic Fungi Isolated from the *Calotropis* procera - A Pharmaceutically Important Xerophytic Plant. International Journal of Research in Pharmaceutical Sciences, 9(1), 82. https://doi.org/10.26452/ijrps.v9i1.1188
- Rashed, K.N.Z. (2024). Medical Plants and Natural Compounds as Anti-Viral Agents: Present and Future Trends. Archives of Biological and Life Sciences, 1(1), 1-25.
- Raunsai, M., Wulansari, D., Fathoni, A., Agusta, A. (2018). Antibacterial and Antioxidant Activities of Endophytic Fungi Extracts of Medicinal Plants from Central Sulawesi. *Journal of Applied Pharmaceutical Science*, 8(8), 069-074.
- Renda, G., Gökkaya, İ., Şöhretoğlu, D. (2021). Immunomodulatory Properties of Triterpenes. *Phytochemistry Reviews*, 21(2), 537-563. <u>https://doi.org/10.1007/s11101-021-09785-x</u>
- Ruzieva, D., Gulyamova, T., Nasmetova, S., Mukhammedov, I., Rasulova, G. (2022). Identification of Bioactive Compounds of the Endophytic Fungus Aspergillus egypticus HT166S Inhibiting the Activity of Pancreatic α-Amylase. Turkish Journal of Pharmaceutical Sciences, 19, 630–635.

- Saxena, P., Selvaraj, K., Khare, S.K., Chaudhary, N. (2021). Superoxide Dismutase as Multipotent Therapeutic Antioxidant Enzyme: Role in Human Diseases. *Biotechnology Letters*, 44, 1– 22.
- Senthil Kumar V., Kumaresan S., Tamizh M.M., Hairul Islam M.I., Thirugnanasambantham K. (2019). Anticancer Potential of NF-κB Targeting Apoptotic Molecule "Flavipin" Isolated from Endophytic *Chaetomium globosum. Phytomedicine*, 61, 152830. https://doi.org/10.1016/j.phymed.2019.152830.
- Shaffique S., Khan M.A., Wani S.H. (2022). A Review on the Role of Endophytes and Plant Growth Promoting Rhizobacteria in Mitigating Heat Stress in Plants. *Microorganisms*, 10, 1286. <u>https://doi.org/10.3390/microorganisms1007128</u> <u>6</u>
- Sharaf M.H., Abdelaziz A.M., Kalaba M.H., Radwan A.A., Hashem A.H. (2022). Antimicrobial, Antioxidant, Cytotoxic Activities and Phytochemical Analysis of Fungal Endophytes Isolated from Ocimum basilicum. Applied Biochemistry and Biotechnology, 194(3), 1271– 1289.
- Sharma H., Rai A.K., Dahiya D. (2021). Exploring Endophytes for In Vitro Synthesis of Bioactive Compounds Similar to Metabolites Produced In Vivo by Host Plants. *AIMS Microbiology*, 7, 175–199.

https://doi.org/10.3934/microbiol.2021012

- Shi X-S, Meng L-H, Li X-M, Li X, Wang D-J, Li H-L, Zhou X-W, Wang B-G. (2019). Trichocadinins B-G: Antimicrobial Cadinane Sesquiterpenes from *Trichoderma virens* QA-8, an Endophytic Fungus Obtained from the Medicinal Plant Artemisia argyi. Journal of Natural Products, 82(9), 2470–2476. https://doi.org/10.1021/acs.jnatprod.9b00139
- Singh, V.K., Kumar, A. (2023). Secondary Metabolites from Endophytic Fungi: Production, Methods of Analysis, and Diverse Pharmaceutical Potential. *Symbiosis*. <u>https://doi.org/10.1007/s13199-023-00925-9</u>

4907

- Suleiman.M.M., Ogah I.J., Okobia B., Adeyemi O.A., Olatunji K.T. & Ige I.M. (2016). A Review On Antibacterial Properties Of Extracts From Psidium Spp And Effect Of The Extraction Solvent. *Pakistan Journal of Medicine and Dentistry*, 5(3). <u>https://ojs.zu.edu.pk/pjmd/articl</u> <u>e/view/2788</u>
- Ujam, N. T., Adonu, C. C., Gugu, T. H., Onwusoba, R., Ike, C., Offiah, R. O., & Esimone, C. O. (2022). Assessment of antiplasmodial and immunomodulatory activities of endophytic fungal metabolites from *Azadirachta indica* A. Juss. *African Journal of Microbiology Research*, 16(3), 121-131.
- Ujam, N. T., Ajaghaku, D. L., Okoye, F. B. C., & Esimone, C. O. (2021). Antioxidant and immunosuppressive activities of extracts of endophytic fungi isolated from *Psidium guajava* and *Newbouldia laevis*. *Phytomedicine Plus*, 1, 100028.
- Usman, A. S., Ali, A., Lawal, M., Liman, M. M., Ameh, L. O., & Kaur, G. (2024). Corn silk: An antidiabetic agent in alloxan-induced diabetic rats. *Journal of Pharmacognosy and Phytochemistry*, 13(2), 776-780. <u>https://doi.org/10.22271/phyto.2024.v13.i2f.149</u> 22
- Venegas-Ortega, M. G., Torres-Llanez, M. J., & González-Córdova, A. F. (2019). The choice of media components and growth conditions can significantly impact the production of bioactive peptides and bacteriocins. *Frontiers in Microbiology*, 10, 965. https://doi.org/10.3389/fmicb.2019.00965
- Wang, W.-X., Zheng, M.-J., Li, J., Feng, T., Li, Z.-H., Huang, R., Zheng, Y.-S., Sun, H., Ai, H.-L., & Liu, J.-K. (2019). Cytotoxic polyketides from endophytic fungus *Phoma bellidis* harbored in *Ttricyrtis maculate*. *Phytochemistry Letters*, 29, 41-46.

https://doi.org/10.1016/j.phytol.2018.11.012

Wu, H., Yan, Z., Deng, Y., Wu, Z., Xu, X., Li, X., Zhou,X., & Luo, H. (2020). Endophytic fungi from the root tubers of medicinal plant *Stephania*

dielsiana and their antimicrobial activity. Acta Ecologica Sinica, 40(5), 383-387.

- Xu, K., Li, X. Q., Zhao, D. L., & Zhang, P. (2021). Antifungal secondary metabolites produced by fungal endophytes: Chemical diversity and potential use in the development of biopesticides. *Frontiers in Microbiology*, 12, 689527. https://doi.org/10.3389/fmicb.2021.689527
- Yamazaki, H., Yagi, A., Takahashi, O., Yamaguchi, Y., Saito, A., & Namikoshi, M. (2020). Antifungal trichothecene sesquiterpenes obtained from the culture broth of marine-derived *Trichoderma cf. brevicompactum* and their structure-activity relationship. *Bioorganic & Medicinal Chemistry Letters*, 30, 127375. https://doi.org/10.1016/j.bmcl.2020.127375
- Yan, Z., Huang, C., Guo, H., Zheng, S., He, J., Lin, J., & Long, Y. (2020). Isobenzofuranone monomer and dimer derivatives from the mangrove endophytic fungus *Epicoccum nigrum* SCNU-F0002 possess α -glucosidase inhibitory and antioxidant activity. *Bioorganic Chemistry*, 94, 103407.

https://doi.org/10.1016/j.bioorg.2020.103407

- Ye, G., Huang, C., Li, J., Chen, T., Tang, J., Liu, W., & Long, Y. (2021). Isolation, structural characterization, and antidiabetic activity of new diketopiperazine alkaloids from mangrove endophytic fungus *Aspergillus* sp. 16-5c. *Marine Drugs*, 19(7), 402. https://doi.org/10.3390/md19070402
- Yehia, R., & Shivanna, M. B. (2021). Antimicrobial activity and metabolite profiling of endophytic fungi in *Digitaria bicornis* (Lam) Roem. and Schult. and *Paspalidium flavidum* (Retz.) A. Camus. 3 Biotech, 11, 53. https://doi.org/10.1007/s13205-020-02587-8
- Yu, F.-X., Li, Z., Chen, Y., Yang, Y.-H., Li, G.-H., & Zhao, P.-J. (2017). Four new steroids from the endophytic fungus *Chaetomium* sp. M453 derived from Chinese Herbal Medicine *Huperzia serrata*. *Fitoterapia*, 117, 41-46. https://doi.org/10.1016/j.fitote.2016.12.012

4908

- Zaden, S. Y. B. D., Qiao, X., & Neefjes, J. (2020). New insights into the activities and toxicities of the old anticancer drug doxorubicin. *FEBS Journal*, 288, 6095-6111. https://doi.org/10.1111/febs.15548
- Zhao, T., Xu, L.-L., Zhang, Y., Lin, Z.-H., Xia, T., Yang, D.-F., Chen, Y.-M., & Yang, X.-L. (2019). Three new α-pyrone derivatives from the plant endophytic fungus *Penicillium ochrochloron* and their antibacterial, antifungal, and cytotoxic activities. *Journal of Asian Natural Products Research*, 21(9), 851-858. <u>https://doi.org/10.1080/10286020.2018.1495197</u>
 - Zhu, J., Lian, J., Wang, X., Wang, R., Pang, X., Xu, B.,& Lu, H. (2023). Role of endogenous and

exogenous antioxidants in the risk of six cancers: Evidence from the Mendelian randomization study. *Frontiers in Pharmacology*, 14. https://doi.org/10.3389/fphar.2023.1185850

Zihad, S. M. N. K., Hasan, M. T., Sultana, M. S., Nath, S., Nahar, L., Rashid, M. A., Uddin, S. J., Sarker, S. D., & Shilpi, J. A. (2022). Isolation and characterization of antibacterial compounds from *Aspergillus fumigatus*: An endophytic fungus from a mangrove plant of the Sundarbans. *Evidence-Based Complementary and Alternative Medicine*, 2022, 9600079. https://doi.org/10.1155/2022/9600079